

# Examining the Effect of Intracellular *C. neoformans* on the Transcriptome of Classically and Alternatively Activated Macrophages



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### Introduction

The fungal pathogen and etiological agent of cryptococcosis, *Cryptococcus neoformans (Cn)*, is responsible for ~181,000 deaths/year worldwide.



**Figure 1.** Cryptococcus neoformans. India ink-stained images of *C. neoformans* by Dr. Erin McClelland

Macrophages, phagocytic innate immune cells, serve as a first line of defense against *Cn* and strongly influence the outcome of infections. The ability of these cells to destroy ingested *Cn* is dependent on their polarization state.

- The classical (M1) state is highly fungicidal and characterized by inducible nitric oxide synthase (iNos) and pro-inflammatory cytokine expression.
- The alternative (M2) state is largely anti-inflammatory and associated with tissue repair.

These polarization states, which are defined by the differential expression of >1000 genes, serve as temporary phenotypes that provide flexibility of function and can change in response to cytokine signals.

As *Cn* is able to replicate inside macrophages, we hypothesized that it may disrupt the polarization state of host cells to compromise fungicidal activity. We also predicted that these transcriptional changes would differ depending on the initial polarization state.

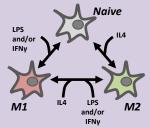
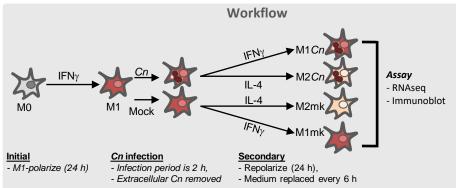


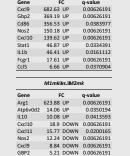
Figure 2. Macrophage polarization. The polarization state of macrophages is inherently plastic and they can be polarized and repolarized in response appropriate to stimuli

## Methods

M1-polarized RAW 264.7 murine macrophages were infected with H99s *Cn.* Post-infection, cells were maintained in M1 or repolarized to M2 using IFN-y or IL-4, respectively. Western blot analysis or RNAseq-based transcriptome profiling of host macrophages was performed 24-hours post-infection. RNAseq was followed by gene ontology using DAVID.



**Figure 3.** Schematic representation of the macrophage infection protocol



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**Table 1.** RNAseq analysis of M1 and M2 macrophage polarization markers

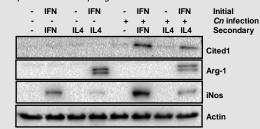
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Figure 4. Intracellular Cn infection affects expression of the M1 marker, iNos at the protein level but not the M2 marker, Arg1. (A) Western blot analysis of protein lysates from cells cultured under the indicated conditions (B) Quantification of iNos and Arg-1 levels by densitometry. Data is from six discrete biological repeats. Error is represented as S.E. \*\* represents p<0.01

# Results (2)

Concordant@p-regulated@enes@n\Cn-infected\ells					Concordant@own-regulated@enes@n\Cn-infected\ells				
	M1mk@s.@M1cn		M2mk@vs.@M2cn			M1mk@s.@M1cn		M2mk@s.@M2cn	
Gene	FC	q-value	FC	q-value	Gene	FC	q-value	FC	q-value
Cited1	14.81	6.26E-03	20.49	1.61E-02	Sorl1	37.12	3.90E-02	3.86	0.046023
Hsf3	7.93	2.86E-02	8.04	0.0200165	Dusp6	14.54	1.29E-02	6.72	0.0375289
Jarid2	5.45	2.52E-02	2.88	0.0389813	C5ar1	12.07	6.26E-03	5.45	0.00626191
Tmtc2	5.14	9.61E-03	3.94	0.00626191	Ehd2	11.63	6.26E-03	5.08	0.00626191
Ccl22	5.09	3.50E-02	4.1	0.00626191	Grk5	8.83	6.26E-03	4.48	0.0128586
Sspn	3.66	4.28E-02	3.21	0.0251778	Cd300ld	7.24	6.26E-03	2.7	0.0389813
Wdr89	3.09	3.34E-02	2.43	0.0389813	Zfp146	6.31	1.61E-02	3.97	0.0304857
Bcl2a1a	2.68	4.68F-02	2.95	0.0171929	Endod1	6.21	6.26F-03	2.57	0.027269

**Table 2.** Identification of genes affected by intracellular Cn infection in both M1 and M2 polarized macrophages



**Figure 5.** The transcriptional co-regulator, Cited1 is up-regulated at the protein level in M1 and M2-polarized host macrophages. Western blot analysis of protein lysates from cells cultured under the indicated conditions. Experiment has been repeated twice.

#### Discussion

Through a combination of RNAseq and western blot analysis, we found that although *Cn* infection had only modest effects on macrophage polarization state (i.e. small increase in the expression of the M1 marker, iNos), it had broader effects on gene expression in host macrophages. We identified a common set of differentially expressed genes that were affected regardless of initial polarization state. This included upregulation of the anti-apoptotic gene, *Bcl2A1a* and the transcriptional coregulator, *Cited1*. Future work will investigate how the expression of these genes influences the fate of host macrophages

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